result from hydrophobic interactions of the Glt- $(\varepsilon Ahx)_{1,2}$ -residues with S_2 and/or other apolar sites on the protein surface. However, this contact does not lead to an acceleration of catalysis due to the failure of the hydrogen bond between CO of P_3 and NH of Gly-216. In the series of Ac-, Glt-, Glt- $(\varepsilon Ahx)_{1,2}$ -Phe-Nan K_M decreases, whereas k_{cat} is nearly constant. Therefore, non-productive binding of the N-acyl residues should not be important, since such interactions lower both K_M and $k_{cat}^{17,18}$. In addition, non-productive binding of the aniline moiety could not be substantiated.

- Abbreviations: Ac-, acetyl-; εAhx-, 6-aminohexanoyl-; Boc-, tert.-butyloxycarbonyl-; Glt-, glutaryl-; Nan-, 4-nitroanilide.
- 2 B.W. Matthews, P.B. Sigler, R. Henderson and D.M. Blow, Nature 214, 652 (1967).
- 3 G.E. Hein and C. Nieman, J. Am. chem. Soc. 84, 4495 (1962).
- 4 M.L. Bender and F.J. Kezdy, A. Rev. Biochem. 34, 49 (1965).
- 5 N. Izumija and T. Yamashita, J. Biochem. 46, 19 (1959).
- 6 C.A. Bauer, C.R. Thompson and E.R. Blout, Biochemistry 15, 1291 (1976).
- 7 M.J. Moffit and G.E. Means, Biochem. biophys. Res. Commun. 83, 1415 (1978).
- 8 D.M. Segal, J.C. Powers, G.H. Cohen, D.R. Davies and P.E. Wilcox, Biochemistry 10, 3728 (1971).
- 9 K. Kurachi, J.C. Powers and P.E. Wilcox, Biochemistry 12, 771 (1973).
- I. Schechter and A. Berger, Biochem. biophys. Res. Commun. 27, 157 (1967).

The high affinity of $Glt-(\varepsilon Ahx)_{1,2}$ -Phe-Nan to the enzyme is reflected in the reaction rate under pseudo 1st-order conditions ($[S] \ll K_M$). Thus, the k_{cat}/K_M value of $Glt-(\varepsilon Ahx)_2$ -Phe-Nan is 3 times higher than that of Glt-Ala-Phe-Nan. In contrast, the k_{cat} values of Glt-Ala-Phe-Nan is 20 times higher than the k_{cat} value of $Glt-(\varepsilon Ahx)_2$ -Phe-Nan.

higher than the k_{cat} value of Glt- $(\varepsilon Ahx)_2$ -Phe-Nan. The k_{cat} and k_{cat} / K_M values in the table show that Glt-Leu-Phe-Nan is more specific than the other substrates studied. Glt-Leu-Phe-Nan should also be of practical interest as a chromophoric chymotrypsin substrate because it is hydrolyzed much faster than the frequently used Glt-Phe-Nan.

- 11 T. Yamashita, J. Biochem. 48, 846 (1960).
- 12 G.L. Neil, C. Nieman and G.E. Hein, Nature 210, 903 (1966).
- M.L. Bender, M.L. Begue-Canton, R.L. Blakeley, L.J. Bru-bacher, J. Feder, C.R. Gunter, F.J. Kezdy, J.V. Killheffer, T.H. Marshall, C.G. Miller, R.W. Roeske and J.K. Stoops, J. Am. chem. Soc. 88, 5890 (1966).
- 14 B.F. Erlanger, F. Edel and A.G. Cooper, Archs. Biochem. Biophys. 115, 206 (1966).
- 15 J. Fischer, L. Lange, S. Koch and H.-D. Jakubke, Eur. J. Biochem. 88, 445 (1978).
- 16 D. Petkov, E. Christova and I. Stoineva, Biochim. biophys. Acta 527, 131 (1978).
- 17 F.E. Brot and M.L. Bender, J. Am. chem. Soc. 91, 7187 (1969).
- 18 J. Fastrez and A. Fersht, Biochemistry 12, 1067 (1973).
- B. Zeeberg, M. Caplow and M. Caswell, J. Am. chem. Soc. 97, 7346 (1975).

C-banding pattern on the chromosomes of the Japanese house shrew, Suncus murinus riukiuanus, and its implication

K. Andō, T. Tagawa and T. A. Uchida^{1,2}

Department of Anatomy, School of Medicine, Fukuoka University, Fukuoka 814 (Japan), and Zoological Laboratory, Faculty of Agriculture, Kyushu University, Fukuoka (Japan), 20 December 1979

Summary. The C-band on the chromosomes of the Japanese house shrew, Suncus murinus riukiuanus (Insectivora), was studied. Various types of C-banding pattern were found in the genome of this subspecies. Such banding patterns could be useful for an understanding of autosome and sex-chromosome polymorphisms within S. murinus.

Karyotypes of the house shrew, Suncus murinus, occurring in various areas of Asia have already been studied, and autosomal and sex chromosomal polymorphisms have been found in this species³⁻⁷, but information on the C-band of the house shrew has as yet only been obtained for the Indian taxon⁷. Nevertheless, C-band staining is a good tool for examining polymorphisms. In this paper, we report the C-banding pattern for the genome of the Japanese house shrew, S.m.riukiuanus, and also present its conventional data again here, because the karyotype figure has not been published in detail.

Materials and methods. 3 male specimens collected from Naha, Okinawa Is., Prefecture of Okinawa, were investigated karyologically. The C-band treatment followed the method of Sumner, and both conventional staining and C-band treatment were performed on the same preparation. For chromosomal classification the method of Patton was adopted.

Results. The C-banding patterns on the autosomes of S.m. riukiuanus are as follows. Within the M · SM-elements (row 1, figure 1), a large pair has a centromeric C-band, which is difficult to detect (see also figure 2A); a small pair and a medium-sized one have a distinct centromeric heterochromatic region; and another small pair exhibits terminal C-bands on its short arm, but does not have centromeric C-band. 2 pairs of ST-autosomes are C-band negative. Of the A-autosomes (row 2 and 3, figure 1), only 5

pairs having a short arm were C-band positive; the remaining 8 pairs seem to be devoid of demonstrable C-band material. 4 pairs of the former A-autosomes have both centromeric C-bands and heterochromatic short arms, but a medium-sized pair (placed in the 2nd position of the 2nd row) is distinguished by a lesser amount of C-banded material than is present in the other 3 pairs. On the other hand, the smallest acrocentric pair (5th in the 2nd row), has a terminal C-band only.

The C-banding pattern on the sex chromosomes (figures 1 and 2, B and C) differs markedly from that of the autosomes. The SM-X chromosome has a broad C-band covering the distal one-third of the long arm and a small terminal C-band on the short arm. The SM-Y chromosome

Lengths of the X and Y chromosomes relative to the female haploid set in the Japanese house shrew, Suncus murinus riukiuanus, calculated from 20 metaphase plates

	X Mean	SD	Y Mean	SD
Relative length	10.49 (5.81)	0.72 (0.35)	5.02	0.60

Values in parentheses show the relative length of only the euchromatic portion of the X chromosome.

has a centromeric C-band and large blocks of interstitial heterochromatin on the long arm. The X and Y chromosomes occupy approximately 10.5% and 5.0% of the female haploid set respectively, and the relative length of the euchromatic portion of the X is about 5.8% of the female haploid set (table).

Discussion. It has been assumed that autosomal polymorphism found in Malaysian and Indian populations of the house shrew originated from a karyotype having 2n = 40 such as that found in S. m. riukiuanus, and that its karyotypic alteration is caused by centric fusions³⁻⁷. The C-banding patterns observed on A-autosomes of the Japanese taxon, as shown in figure 1B, represent rather differentiated figures. Consequently, by their C-banding patterns, the A-autosomal complement can be classified into 2 groups, and some pairs in that can be clearly identified. It would be expected, therefore, that the C-band patterns characterized on A-autosomes of S. m. riukiuanus could serve as material for the discussion about which uniarmed elements gave rise to the additional biarmed ones found in Malaysian and Indian populations. In fact, logical explanations referring to intraspecific karyotype evolution have been made in some rodents using the C-band¹¹⁻¹³.

The C-banding patterns of the sex chromosomes in the Japanese taxon may explain the intraspecific sex chromosome polymorphism noted in the Asian S. murinus. It has been shown, from conventional data, that the biarmed X chromosome of the West Bengal (India) population¹⁴ is evidently smaller than those of other populations in Asia⁵. As for the Y chromosome, in the Indian¹⁴ and South Vietnamese populations¹⁵ the Y has been regarded as the smallest A-element in each complement. On the other hand, in the West Malaysian population⁵, except for the Malacca one, the Y chromosome has been established as

M-element, being larger than those of the above 2 populations. In S. m. riukiuanus the Y is SM-element and appears larger in size than the M-Y of the West Malaysian population. In another Malaysian population (Malacca), the Y chromosome is 6.0-7.5% of the female haploid set⁵, though its morphology resembles that of the Japanese taxon. A larger Y (7.7-8.6% of the female haploid set) has been found in the Indian population from Delhi and Varanasi^{16,17}. The large X chromosome in S. murinus is in agreement with the theory of the duplicate type X discussed by some workers^{18,19}, because the X chromosomes of S. m. riukiuanus and the Indian taxon7 possess a high amount of Cband material. This assumption is supported also by the facts that the euchromatic regions of the X chromosome in the Japanese taxon is approximately equal to the size of a typical eutherian X, and that autosomal Robertsonian translocation found in Malaysian and Indian has no effect on the formation of large X (no X-autosomal translocation). Thus, size variation of the X chromosome in the house shrew seems to be attributable to differences in the amount of C-band material.

Similarly, the long Y chromosome of the Japanese house shrew also contains a large amount of heterochromatin, although it is not C-band material alone. The amount and location of C-bands on the Y chromosomes of most mammals are variable 13,19,20. Further, in Australian rodents, the Y chromosome polymorphism is due to the variation of constitutive heterochromatin 21,22. Given the data presented in previous studies and the C-banding pattern on the Y chromosome, we believe that differences in the amount of constitutive heterochromatin are a chief factor in variation regarding shape and size of the Y chromosome in S. murinus. The prototypic Y in this species was probably a small acrocentric.

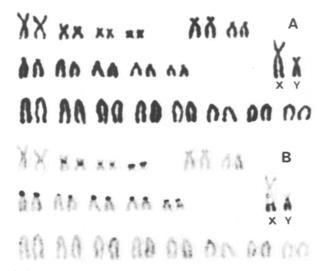


Fig. 1. Conventional (A) and C-banded (B) karyotype of the same cell in the Japanese house shrew, *Suncus murinus riukiuanus*.

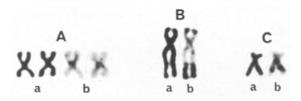


Fig. 2. Standard conventional features (a) and their C-banding patterns (b) of a large metacentric or submetacentric autosome pair (A), the X chromosome (B) and the Y chromosome (C).

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- 2 Zoological Laboratory, Faculty of Agriculture, Kyushu University, Fukuoka (Japan).
- 3 H.S. Yong, Experientia 15, 589 (1971).
- 4 H.S. Yong, Experientia 15, 585 (1972).
- 5 H.S. Yong, Caryologia 27, 65 (1974).
- 6 N.V. Aswathanarayana and K.L. Satya Prakash, in: Chromosomes today, vol.5, p.447. Ed. P.L. Pearson and L.R. Lewis. Wiley, New York 1974.
- 7 K. L. Satya Prakash and N.V. Aswathanarayana, Proc. Dunn Dobzh. Symp. Genet. 272 (1976).
- 8 T.A. Uchida and K. Andō, Sci. Bull. Fac. Agric. Kyushu Univ. 26, 393 (1972).
- 9 A. T. Sumner, Exp. Cell Res. 75, 304 (1972).
- 10 J.L. Patton, J. Mammal. 48, 27 (1967).
- 11 J.T. Mascarello and J.W. Warner, Experientia 30, 90 (1974).
- 12 T.H. Yosida and T. Sagai, Chromosoma 50, 283 (1975).
- 13 P. R. Berverstock, C.H.S. Watts and J.T. Hogarth, Chromosoma 61, 95 (1977).
- 14 G.K. Manna and M. Talukdar, Mammalia 31, 288 (1967).
- 15 J.F. Duncan, P.F.D. Van Peenen and P.F. Ryan, Caryologia 23, 173 (1970).
- 16 S.R.V. Rao, V.K. Sharma and V.C. Shah, Cytogenetics 9, 384 (1970).
- T. Sharma, S. Pathak and S.P. Ray-Chaudhuri, The Nucleus 13, 62 (1970).
- 18 S. Ohno, W. Beçak and M.L. Beçak, Chromosoma 15, 14 (1964).
- 19 T.C. Hsu and F.E. Arrighi, Chromosoma 34, 243 (1971).
- T.R. Chen and F.H. Ruddle, Chromosoma 34, 51 (1971).
 W.H. Bradshaw and T.C. Hsu, Cytogenetics 11, 436 (197).
- W.H. Bradshaw and T.C. Hsu, Cytogenetics 11, 436 (1972).
 P.R. Bayerstock, C.H.S. Watts and J.T. Hogarth, Chromosomer Chromos
- P. R. Baverstock, C.H.S. Watts and J.T. Hogarth, Chromosoma 61, 243 (1977).